



Dissociation Between Phosphodiesterase Inhibition and Antiproliferative Effects of Phosphodiesterase Inhibitors on the Dami Cell Line

Katja Zurbonsen,^{*||} Alain Michel,[†] Daniel Vittet,[‡]
Pierre-Antoine Bonnet[§] and Claude Chevillard^{*}

^{*}INSERM U.300, [†]LAB. PHARMACODYNAMIE, [‡]INSERM U.217 AND [§]LAB. DE CHIMIE ORGANIQUE
PHARMACEUTIQUE, FACULTE DE PHARMACIE, 15, AV. CHARLES FLAHAUT, 34060 MONTPELLIER CEDEX 2, FRANCE

ABSTRACT. Phosphodiesterase (PDE) inhibitors were shown to inhibit proliferation of various cell types. The present investigation was designed to study the activity of selective PDE inhibitors (8-MeoMIX, milrinone, trequinsin, rolipram, RO-201724, zaprinast, and MY-5445) on the proliferation of the Dami cell line in relation to their effects on cAMP levels and PDE isoenzymes isolated from Dami cells. All compounds, except 8-MeoMIX, elicited antiproliferative effects. Trequinsin, RO-201724, and MY-5445 (100 μ M) were found to inhibit cell growth up to 60%, 83%, and 85%, respectively; milrinone, rolipram and zaprinast elicited only weak effects (19–21% at 100 μ M). Their growth-inhibitory effects could not be related to their effects on cAMP levels. In addition, although PDE type III and IV inhibitors potentiated cAMP formation due to adenylcyclase activation, no potentiation could be observed when considering their antiproliferative effect. Separation and characterization of PDE of Dami cells revealed the existence of types III, IV, and V isoenzymes. The PDE inhibition found for the PDE inhibitors could not explain their antiproliferative effects. The lack of correlation with cAMP concentrations or PDE inhibition and the high concentrations needed to elicit antiproliferative effects suggest the implication of other parameters, such as cytotoxicity or lipophilicity, or other targets in addition to PDE for the PDE inhibitors tested. Lipophilicity did not seem to be of importance in antiproliferative effects. In contrast, cytotoxic effects, in particular those of trequinsin and MY-5445, could partially explain their negative action on cell growth. *BIOCHEM PHARMACOL* 53;8:1141–1147, 1997. © 1997 Elsevier Science Inc.

KEY WORDS. Dami cell line; PDE inhibitors; PDE isoenzymes; proliferation; cAMP; megakaryoblast

cAMP[¶] is known to be involved in the proliferation of different cell types. It can be a positive intracellular signal for cell growth, as shown for many differentiated cells [1, 2] as well as a negative messenger as described for various cancer cells [2–5]. Rises in intracellular cAMP levels can be induced by inhibitors of the cAMP-degrading enzymes, the PDE. Previously, studies have shown that nonselective PDE inhibitors of the methylxanthine type [3, 5–10], pyrimido-pyrimidines [11–13] and imidazo[1,2-a]pyrazine derivatives [14] could display antiproliferative effects on normal and cancer cell lines. At present, little is known about the activity of selective PDE inhibitors [15–17] on cell proliferation. Studies have been carried out with different selective-type I, III, IV, V, or mixed-type III/IV PDE inhibitors on various cell lines [18–23]. All PDE inhibitors of type III

[19–23] as well as those of type IV [18, 19, 21–23] and of mixed type III/IV [19, 20] were shown to inhibit cell growth of the different cell types tested. These antiproliferative activities have been correlated to intracellular cAMP levels and to the PDE inhibitory potency of these compounds [19–21, 23].

In the present study, we have investigated the effects of a series of selective PDE inhibitors (Table 1) on the proliferation of the human megakaryoblastic leukemic Dami cell line [24], which was found to be negatively regulated by cAMP [5], in relation to their ability to increase intracellular cAMP levels and their PDE inhibitory potency toward isoenzymes isolated from Dami cells.

MATERIALS AND METHODS

Materials

Dami cells were obtained from the American type culture collection (CRL 9792) (ATCC, Rockville, MD). Horse serum was purchased from Techgen International (Les Ulis, France), and cell culture medium and PBS Dulbecco's were from Gibco BRL (Eragny, France).

Trequinsin, milrinone, RO-201724, rolipram, zaprinast

^{||} Corresponding author. Katja Zurbonsen, INSERM, U.300, Faculté de Pharmacie, 15, Av. Charles Flahaut, 34060 Montpellier Cedex 2, France. Tél. 04 67 52 46 92; FAX 04 67 04 21 40.

[¶] Abbreviations: PDE, phosphodiesterase; cAMP, cyclic 3', 5'-adenosine monophosphate; cGMP, guanosine 3',5'-cyclic monophosphate; PKA, cAMP-dependent protein kinase; LDH, lactate dehydrogenase, PGE1, prostaglandin E1.

Received 24 June 1996; accepted 8 October 1996.

TABLE 1. Selective PDE inhibitors used

PDE inhibitor*	Type
8-MeoMIX	I
Milrinone, trequinsin	III
Rolipram, RO-201724	IV
MY-5445, zaprinast	V

* Classification as proposed by Beavo (17).

and MY-5445 were purchased from Biomol (Tebu, le Perray en Yvelines, France) and 8-MeoMIX from LC-laboratories (Meudon, France-Biochem, France). The cAMP 125 I-radioimmunoassay kit was from Immunotech (Marseilles, France) and the CytoTox 96 kit from Promega (Charbonnières, France). Radiochemicals were obtained from Amersham (Les Ulis, France). Other reagents were purchased from Sigma (St. Quentin Fallavier, France).

Cells and Cell Culture Conditions

Cells were grown in suspension in RPMI 1640 medium supplemented with 10% heat-inactivated horse serum, 2 mM glutamine, 1 mM sodium pyruvate, and 0.4% nonessential amino acids. They were incubated in disposable sterile Erlenmeyer flasks (Corning, Polylabo, Strasbourg, France) at 37°C with a humidified atmosphere of 5% CO₂. Cultures were fed every 2–3 days by partial replacement of spent medium.

Proliferation Studies

Cellular proliferation was studied by placing cells in the exponential phase of growth in microwells (Nunc, Polylabo, Strasbourg, France) at a starting concentration of 2×10^5 cells/mL. Culture conditions were as described above. Cells were incubated with the products to be tested for 48 hr and cell proliferation was determined by cell counting performed using a Coulter Counter ZM (Coultronics S. A., Margency, France) equipped with a 140- μ m orifice tube, and calibrated with 14 μ m latex particles.

Cell Viability

Cell viability was determined by the use of the CytoTox 96 nonradioactive assay, which allows the quantitative spectrophotometric measurement of LDH activity [25]. Cells were seeded and incubated with the different compounds for 48 hr as described in Vittet et al. [5].

Cyclic AMP Assays

Samples for determination of intracellular cAMP concentrations by a radioimmunoassay kit were prepared as follows. Cells were washed with PBS (158 mM NaCl; 2.7 mM KCl; 1.5 mM KH₂PO₄; 8 mM Na₂HPO₄; 0.5 mM MgCl₂; 0.9 mM CaCl₂; 5.55 mM glucose, pH 7.4), resuspended to

a final concentration of 2.0×10^6 cells/mL and preincubated for 5 min at 30°C before addition of compounds to be tested. Incubations were performed at 30°C for 10 min and terminated by adding 60% cold trichloroacetic acid to a final concentration of 10%. After mixing, each sample was spun at 14,000 g for 2 min and the supernatants were neutralized with KOH (3 mM) and buffered with HEPES (150 mM). The suspension was kept in an ice bath for 10 min and the precipitate was removed by centrifugation at 2500 g for 5 min. The neutralized extracts were stored at –20°C. cAMP concentration was assayed according to the manufacturers' recommendations.

Preparation of Cell Extracts and High-Speed Supernatant

Cells were separated from culture medium by centrifugation at 150 g for 5 min. After 2 washes with PBS Dulbecco's buffer without calcium, magnesium, or sodium bicarbonate, the cell pellet (10^8 cells) was resuspended in 2 mL extraction buffer (10 mM Tris-HCl, 2 mM MgCl₂, 10 μ M Antipain, 10 μ M Leupeptin, 10 μ M Pepstatin A, 5 mM β -mercaptoethanol, 200 μ M phenylmethylsulfonyl fluoride, pH 7.5), homogenized, and centrifuged at 1000 g for 15 min. The supernatants were centrifuged at 100,000 g for 60 min to obtain the high-speed supernatant.

Separation of PDE Isoenzymes

PDE isoenzymes were separated from the high-speed supernatant by anion exchange chromatography. The supernatant (1600 μ g protein) was applied to a DEAE-Sepharose CL-6B column (Sigma, St. Quentin Fallavier, France) pre-equilibrated with the extraction buffer (described above) containing 0.1 M Na acetate. PDE isoenzymes were successively eluted by using a step-by-step Na acetate gradient in the same buffer (see Fig. 4) and collected as 1 mL fractions. For storage at –20°C, ethylene glycol was added to a final concentration of 30% (v/v).

Protein concentration was determined as described by Lowry et al. [26].

PDE Assay

The PDE activity of cell extracts was determined by the method of Thompson et al. [27], modified by Cook et al. [28]. Assays were performed at 37°C in a total volume of 100 μ L. Each tube contained 25 μ L of each fraction, 50 μ L of assay buffer (final concentration of 40 mM Tris-HCl, 2.5 mM MgCl₂, 3.75 mM β -mercaptoethanol, 0.2 μ Ci [3 H]-cAMP or cGMP, 1 μ M cAMP, or cGMP, pH 8.0) and 25 μ L PDE inhibitor or its respective solvent. Following 30 min incubation, the reaction was stopped by transferring to a bath of boiling water for 3 min. After cooling on ice, 20 μ L of 1 mg/mL *Ophiophagus hannah* venom was added and the reaction mixture was incubated at 37°C for 10 min. Unreacted [3 H]-cAMP or [3 H]-cGMP was removed by the

addition of 400 μL of a 35% suspension of Dowex 1 \times 8–400 μL resin and incubation on ice for 30 min. After centrifugation (2500 g, 5 min), 200 μL of the supernatant was removed for liquid scintillation counting. Less than 10% of the tritiated cyclic nucleotide was hydrolyzed in any assay.

The concentration of the PDE inhibitors that causes 50% PDE inhibition (IC_{50}) was determined from concentration-response curves. At least two concentration-response curves were generated for each agent.

Statistical Evaluation

Results are means \pm SE. Statistical differences were appreciated by Student's *t*-test or Withney-U test (cAMP assay). $P < 0.05$ was considered as significant. IC_{50} values were calculated using computer software [29] and lipophilicity values ($\log P$) were evaluated as described by Viswanadhan et al. [30].

RESULTS

Effects of the PDE

Inhibitors on Dami Cell Proliferation

None of the compounds affected cell growth at concentrations below 1 μM (1–100 nM). RO-201724, trequinsin, and MY-5445 induced a potent concentration-dependent cell growth inhibition at the highest concentration assayed (100 μM) up to 60%, 83%, and 85%, respectively (Fig. 1, Table 2). For milrinone, rolipram and zaprinast, only weak

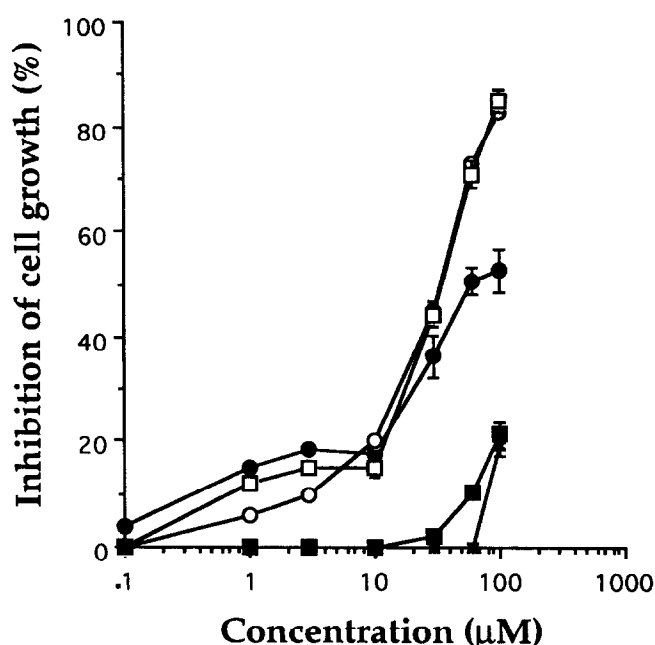


FIG. 1. Concentration-dependent activity of RO-201724 (●) trequinsin (○), MY5445 (□), rolipram (■) and Milrinone (×) on Dami cell growth. Growth inhibition is expressed in percent related to control. Each point represents the mean \pm SE of 3 experiments performed in duplicate. Each value of duplicate is based on 4 cell counts.

significant growth inhibitions of $19.2\% \pm 1.2$, $21.5\% \pm 2.8$ and $21.2\% \pm 1.9$, respectively, were found at the highest concentration (100 μM) tested. 8-MeoMIX was devoid of any effect. Due to the poor compound solubility, 100 μM could not be exceeded.

Determination of Cytotoxicity

We observed a significant increase in LDH release from cells induced by trequinsin and MY-5445 at 10 μM and above (Fig. 2A, B). A correlation ($R = 0.93$) between LDH release and inhibition of cell proliferation was found for these compounds. 100 μM rolipram also presented significant cytotoxic activity (14.8 ± 1.3 vs 8.5 ± 0.8), whereas neither 100 μM RO-201724, milrinone, nor zaprinast significantly increased LDH release (results not shown).

Effects of the PDE Inhibitors on Intracellular cAMP Levels in the Presence and Absence of Prostaglandin E1 (PGE1)

To look for a potential relationship between cell growth inhibition and intracellular cAMP concentrations, we investigated the effect of the compounds on cAMP concentration in the presence and absence of the adenylyl cyclase stimulator PGE1. PGE1 was tested at 0.1 μM , a concentration that induced only a low cAMP increase in Dami cells (37.7 ± 11.4 pmol/million cells). In the absence of PGE1, only milrinone and trequinsin significantly increased cAMP concentration (Fig. 3A). All compounds tested, except 8-MeoMIX, MY-5445, and zaprinast, potentiated cAMP formation in the presence of PGE1 (Fig. 3B).

Effects of Milrinone and RO-201724 on Dami Cell Growth in the Presence and Absence of PGE1

To investigate whether or not the antiproliferative effects of milrinone and RO-201724 were potentiated in the presence of adenylyl cyclase stimulation, we compared their activity on cell growth after 48 hr incubation at 100 μM and 10 μM (only RO-201724) in the presence and absence of PGE1 (Table 3). PGE1 (0.1 μM) elicited weak proper antiproliferative effects ($11.6\% \pm 0.5$) and did not potentiate the antiproliferative effect of 100 μM Milrinone or RO-201724 (10 and 100 μM). Only additive effects on cell growth inhibition were observed for these compounds. Furthermore, we did not observe any potentiation of the effect of RO-201724 at 0.1 and 1 μM in the presence of PGE1 (data not shown).

Distribution of Cyclic Nucleotide PDE Activity in Dami Cells

Dami cells contained both cAMP PDE and cGMP PDE activities. Over 85% of total cAMP and 85% of total cGMP PDE activity was found in the cytosolic fraction.

TABLE 2. Effects of PDE inhibitors on PDE isoenzymes from Dami cytosol and on cell growth inhibition (G.I.)

Compounds†	log P	-log (IC ₅₀)*			
		G.I.	Type III (peak 3)	Type IV (peak 2)	Type V (peak 1a/b)
Trequinsin	1.97	4.6 ± 0.1	8.2 ± 0.2	5.8 ± 0.3	N.D.‡
Milrinone	-0.57	<4.0	5.2 ± 0.1	4.7 ± 0.1	N.D.‡
Rolipram	1.66	<4.0	<4.0	5.2 ± 0.1	N.D.‡
RO-201724	1.81	4.3 ± 0.0	<4.0	4.1 ± 0.2	N.D.‡
Zaprinast	1.36	<4.0	N.D.‡	N.D.‡	5.4 ± 0.1
MY-5445	6.58	4.5 ± 0.0	N.D.‡	N.D.‡	<5.0

* Values are the means ± SE of N = 7–10 experiments performed in triplicate, † Compounds were assayed in the presence of 1 μM cAMP or cGMP as substrate ‡ ND, not determined.

Characterization of the PDE Isoforms of Dami Cytosol

Four peaks of cyclic nucleotide PDE activity were separated by Na acetate gradient elution of the cytosolic fraction (Fig. 4). Peaks 1a and 1b revealed specific hydrolytic activity for cGMP. This activity was not stimulated by Ca²⁺-calmoduline (results not shown) and was inhibited by zaprinast and MY-5445 (Table 2). According to Beavo and Reifsnnyder [17, Table 1], these characteristics apply to PDE type V. The isoenzymes eluted in peaks 2 and 3 preferentially hydrolyzed cAMP. The isoenzymes eluted in peak 2 were insensitive to cGMP (10 μM), not stimulated by Ca²⁺-calmoduline (results not shown), and were primarily inhibited by rolipram and RO-201724 (Table 1). These are specific characteristics for type IV isoenzymes. The specific PDE type III inhibitors milrinone and trequinsin showed preferences for the isoenzymes of peak 3 (Table 2), which were also inhibited by cGMP (38.1% ± 0.6 at 10 μM; 52.4% ± 3.3 at 1 μM). The trequinsin IC₅₀ for the cAMP hydrolytic activity of peak 3 was 1000 times lower than that of peak 2. These characteristics apply to PDE type III.

DISCUSSION

The PDE type I and V inhibitors tested in our study presented only marginal (zaprinast) or undetectable (8-Meomix) effects on Dami cell proliferation at the highest concentration tested, which was consistent with studies from other groups [18, 23]. Cell growth inhibition elicited by MY-5445 appears essentially related to its cytotoxic effect.

PDE inhibitors type III and IV have shown antiproliferative effects on Dami cell proliferation. Among them, trequinsin and RO-201724 inhibited cell growth in a concentration-dependent manner. Milrinone and rolipram presented only a weak antiproliferative effect at the highest concentrations tested. These results are in agreement with other studies that have investigated the activity of different selective type III, IV, or mixed type III/IV PDE inhibitors on proliferation of lymphocytes [18, 19], smooth muscle cells [20–22], B16 murine melanoma and human MCF-7 mammary carcinoma cell lines [23]. In these studies, all PDE inhibitors of type III, SK&F 94836 [19, 21], milrinone [18], trequinsin [21], CI-930, and cilostamide [22], as well as

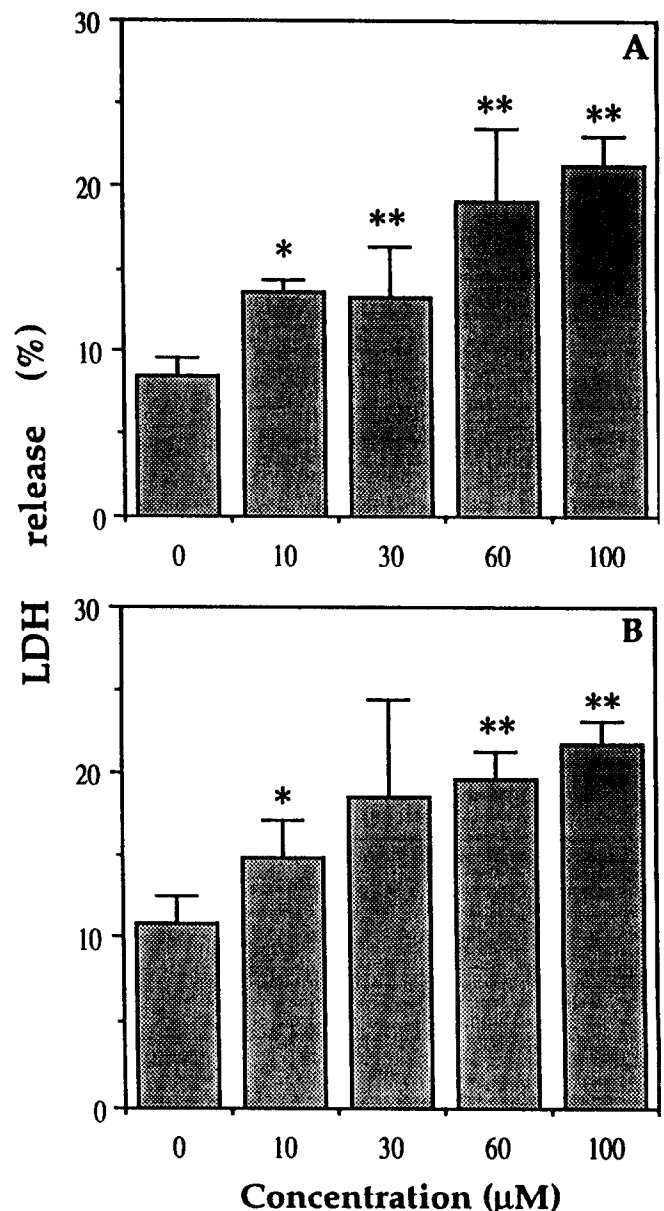


FIG. 2. Concentration-dependent activity of (A) Trequinsin, (B) MY5445, and their respective controls on LDH release from Dami cells after 48-hr incubation. Histograms are the means and vertical bars indicate the SE of at least 4 experiments performed in triplicate. Significantly different from controls: ***P* < 0.01, **P* < 0.05.

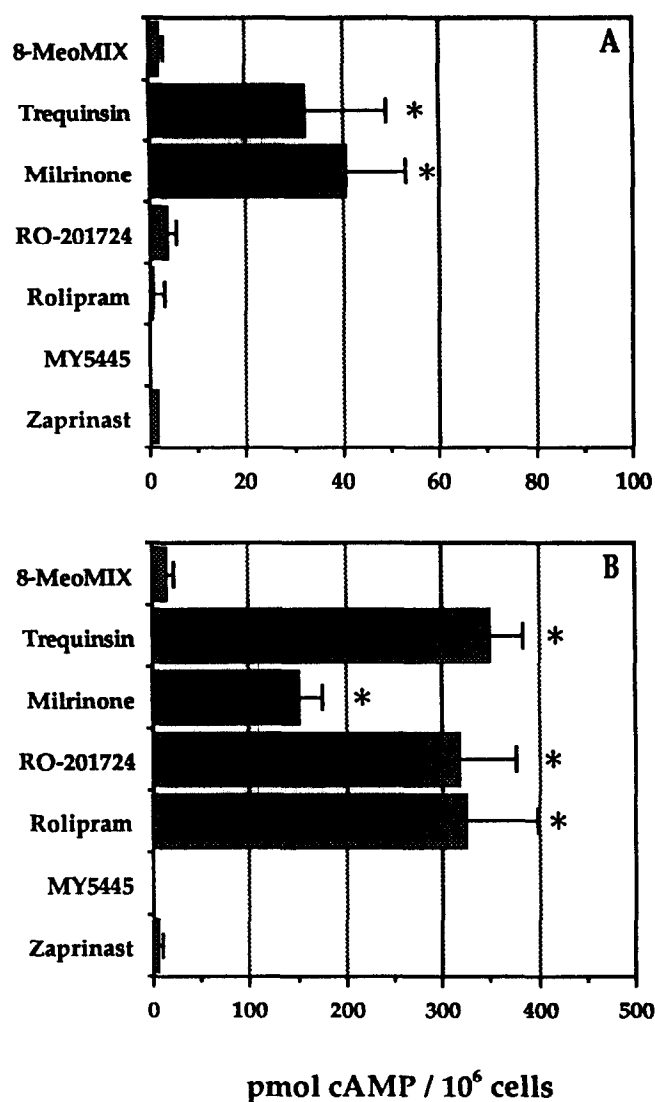


FIG. 3. Effects of 100 μ M of the PDE inhibitors on intracellular cAMP levels (A) in absence, and (B) in presence of 0.1 μ M PGE1. Histograms are the means and horizontal bars indicate the SE of 3 to 5 experiments performed in duplicate. * $P < 0.01$.

those of type IV, DC-TA-46 [23], rolipram, and RO-201724 [18, 19, 21, 22] and those of mixed type III/IV, zardaverine and cilostazol [19, 20], were shown to inhibit cell growth.

Although most PDE inhibitors tested here elicited cell-growth inhibition, very high concentrations up to 100 μ M were needed to induce that effect. Moreover, such high concentrations of selective PDE inhibitors have already been described to be necessary to induce cell-growth inhibition [18, 21, 22].

In view of the high concentrations needed, it appeared difficult to explain this antiproliferative activity by their PDE-inhibitory potency. To clarify this point, we investigated if the antiproliferative activities of the PDE inhibitors were related to their effects on intracellular cAMP. Our results indicated no correlation between these two param-

TABLE 3. Effects of selective PDE inhibitors on Dami cell growth

Compounds	G.I. (%) [*]	
	In absence of PGE1	In presence of PGE1 (0.1 μ M)
PGE1	—	11.6 \pm 0.5
Milrinone [†]	18.7 \pm 0.7	34.3 \pm 3.3
RO-201724 [†]	39.5 \pm 7.9	57.2 \pm 2.4
RO-201724 [‡]	24.9 \pm 2.0	42.8 \pm 2.8

^{*} G.I., growth inhibition, values are means \pm SE, N = 4–8.

[†] 100 μ M.

[‡] 10 μ M.

eters. Indeed, rolipram and RO-201724 inhibited cell growth at the highest concentration tested without affecting intracellular cAMP levels. In contrast, trequinsin and milrinone induced intracellular cAMP levels in the same manner, but exhibiting quite different effects on cell proliferation (i.e. marginal for milrinone and potent for trequinsin). In addition, although the effects of the PDE type III and IV inhibitors on intracellular cAMP levels were potentiated in the presence of PGE1, their antiproliferative activity was only added to those of PGE1, which confirms a lack of correlation between cell-growth inhibition and intracellular cAMP levels.

The need to verify a potential relationship between antiproliferative activity and PDE inhibitory potency led us to isolate the PDE isoenzymes from Dami cells.

Separation of these isoenzymes revealed the presence of a peak of cGMP-specific hydrolyzing isoenzymes, charac-

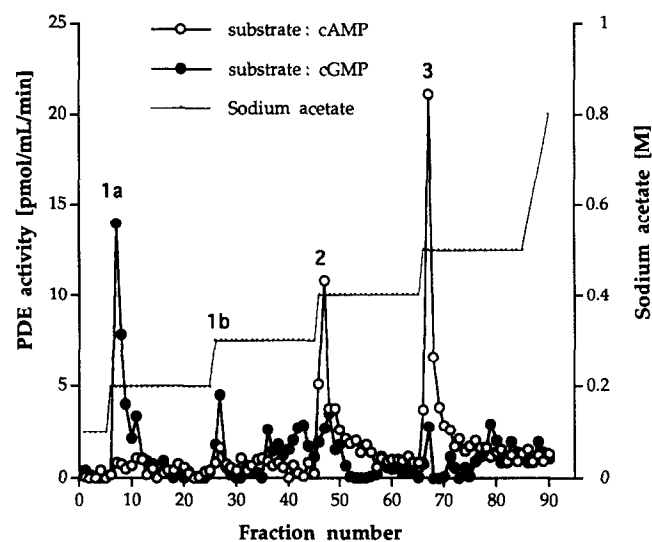


FIG. 4. DEAE-Sepharose-CL-6B anion exchange chromatography of PDE activity of Dami cytosol: cAMP-PDE activity was assayed with 1 μ M cAMP (\circ) and cGMP-PDE activity with 1 μ M cGMP (\bullet) as substrate. The shape of the Na acetate gradient is shown. Fractions of 1 mL were collected and 30% ethylene glycol (v/v) was added. The activity yield of the 90 fractions collected represents 80% of total cAMP-PDE and 90% of total cGMP-PDE.

terized by the inhibitory effects of zaprinast and MY-5445 as PDE type V, as well as the presence of 2 peaks of cAMP-specific hydrolyzing isoenzymes. In view of the inhibitory effects of the type III and IV inhibitors, we characterized these isoenzymes as type III and IV. These results were confirmed by the observation that cGMP selectively inhibited PDE isoenzymes characterized as type III, whereas it presented no effect on those of type IV.

It is noteworthy that only rolipram—trequinsin elicited a good PDE III–PDE IV discriminating potency, RO-201724—milrinone being less selective. However, milrinone presented the same inhibitory potency on the PDE III isoenzymes when used in the presence of 10 μ M rolipram (results not shown). All these observations show a satisfactory separation of type III and IV isoenzymes. However, the IC_{50} values observed here were 6 to 36 times higher than those described for other systems [23, 31–36]. Both high IC_{50} values and discriminating potencies of RO-201724 and milrinone revealed quite unusual reactivities for the PDE isoenzymes isolated from Dami cells that may be related to the existence of different genetic subtypes, as has already been described for type III and IV isoenzymes [37]. When comparing IC_{50} values for PDE and growth inhibitions, there appears at least a 2-log difference and no correlation between the two parameters. Indeed, RO-201724 elicited equipotent antiproliferative effects in the 1–10 μ M range, and its PDE inhibitory potency varied from 0 to 38.5%. Furthermore, rolipram and milrinone did not affect cell growth at lower concentrations (from 1 to 10 μ M), but presented PDE inhibitory effects from 30 to 58.6% and from 17.8 to 62.5% in the same concentration range, respectively. In addition, trequinsin inhibited PDE type III at nM concentration (IC_{50} = 6 nM) without affecting cell proliferation at these concentrations. Studies from other authors have also shown discrepancies between the PDE and growth IC_{50} values [18, 21, 22] of PDE inhibitors, although their growth-inhibitory effect has been related to their PDE-inhibitory potency.

In summary, it appears that the effects of the selective PDE inhibitors tested on Dami PDE isoenzyme activity and on intracellular cAMP levels could not totally explain proliferation responses. These results are in contrast to other studies realized with selective PDE inhibitors. The antiproliferative effect of the type IV inhibitor DC-TA-46 has been correlated to intracellular cAMP levels and to its PDE-inhibitory potency [23]. Furthermore, for the type III inhibitor SK&F 94836 and type IV inhibitor rolipram, alone or in combination, as well as for cilostazol and zardaverine, increased cAMP levels were reported to mediate the inhibitory effect on cell growth [19–21]. This discrepancy between our results and those found previously suggests the implication of other parameters, in addition to PDE inhibition, for the PDE inhibitors tested, such as cytotoxicity and differences in lipophilicity, and targets other than PDEs. The potent antiproliferative effects of trequinsin and MY-5445 seem to be partially related to their cy-

totoxic effects. Although high concentrations of PDE inhibitors have been used in some studies to induce potent effects on cell growth inhibition [18, 22], no such cytotoxic effects have been reported. Evaluation of the lipophilicity values of the PDE inhibitors has indicated an absence of linkage between this parameter and cell-growth inhibitory effects.

PDE inhibitors have also been reported to have other targets in addition to PDEs, in particular, cAMP-dependent protein kinase (PKA) [38] or Gi protein [39]. Furthermore, there is evidence for a “cross-talk” between PDEs and other signalling systems that could regulate PDE activity and/or expression [40]. It has been suggested that such interactions or other targets modulate the PDE inhibitory activity of RO-201724, MY-5445 and zaprinast [41–43].

This study shows that some of the selective PDE inhibitors tested exhibited inhibitory effects on the growth of the Dami cancer cell line. However, in view of the high concentrations needed to elicit cell-growth inhibition and the absence of any correlation with intracellular cAMP levels or selective PDE-inhibitory potency, it appears that side effects of the PDE inhibitors also participate in Dami cell-growth inhibition.

The authors address special thanks to Drs. F. et M. F. Laliberté for their helpful suggestions. The authors are grateful to Dr. S. M. Greenberg for providing Dami cells.

References

1. Dumont JE, Jauniaux JC and Roger PP, The cyclic AMP-mediated stimulation of cell proliferation. *Trends Biochem Sci* 14: 67–71, 1989.
2. Boynton AL and Whitfield JF, The role of cyclic AMP in cell proliferation: a critical assessment of the evidence. *Adv Cyclic Nucleotide Res* 15: 193–294, 1983.
3. Cho-Chung YS, Role of cyclic AMP receptor proteins in growth, differentiation and suppression of malignancy: new approaches to therapy. *Cancer Res* 50: 7093–7100, 1990.
4. Cho-Chung YS, Clair T, Tortora G and Yokozaki H, Role of site-selective cAMP analogs in the control and reversal of malignancy. *Pharmacol Ther* 50: 1–33, 1991.
5. Vittet D, Duperray C and Chevillard C, Cyclic-AMP inhibits cell growth and negatively interacts with platelet membrane glycoprotein expression on the Dami human megakaryoblastic cell line. *J Cell Physiol* 163: 645–655, 1995.
6. Leitman DC, Fiscus RP and Murad F, Forskolin, phosphodiesterase inhibitors, and cyclic AMP analogs inhibit proliferation of cultured bovine aortic endothelial cells. *J Cell Physiol* 127: 237–243, 1986.
7. Lando M, Abemayor E, Verity MA and Sidell N, Modulation of intracellular cyclic adenosine monophosphate levels and the differentiation response of human neuroblastoma cells. *Cancer Res* 50: 722–727, 1990.
8. Prasad KN and Sheppard JR, Inhibitors of cyclic nucleotide phosphodiesterase induce morphological differentiation of mouse neuroblastoma cell culture. *Exp Cell Res* 73: 436–440, 1972.
9. Janik P, Assaf A and Bertram JS, Inhibition of growth of primary and metastatic Lewis lung carcinoma cells by the phosphodiesterase inhibitor isobutylmethylxanthine. *Cancer Res* 40: 1950–1954, 1980.
10. Fontana JA, Miksis G, Miranda DM and Durham JP, Inhibi-

- tion of human mammary carcinoma cell proliferation by retinoids and intracellular cAMP-elevating compounds. *J Natl Cancer Inst* **78**: 1107–1112, 1987.
11. Biddle W, Montagna RA, Leong SS, Horoszewicz J, Gastpar H and Ambrus JL, Antineoplastic effect of the pyrimido-pyrimidine derivative: RA 233. *Pathol Biol* **32**: 9–13, 1984.
 12. Lichtner RB, Goka TJ, Butcher RW and Nicolson GL, Direct effects of the pyrimido-pyrimidine derivative RA 233 (Rap-enton) on rat 13762NF mammary tumor cell clones *in vitro*. *Cancer Res* **47**: 1870–1877, 1987.
 13. Lichtner RB and Nicolson GL, The pyrimido-pyrimidine derivatives RA 233 and RX-RA 85 affect growth and cytoskeletal organization of rat mammary adenocarcinoma cells. *Eur J Cancer Clin Oncol* **23**: 1269–1275, 1987.
 14. Levallois C, Bonnafous JC, Françoise MA, Sablayrolles C, Chapat JP and Mani JC, Theophylline-like properties of xanthine analogs. *Biochem Pharmacol* **33**: 2253–2257, 1984.
 15. Beavo JA, Conti M and Heasley RJ, Multiple cyclic nucleotide phosphodiesterases. *Mol Pharmacol* **46**: 399–405, 1994.
 16. Nicholson CD, Challiss RAJ and Shahid M, Differential modulation of tissue function and therapeutic potential of selective inhibitors of cyclic nucleotide phosphodiesterase isoenzymes. *Trends Pharmacol Sci* **12**: 19–27, 1991.
 17. Beavo JA and Reifsnnyder DH, Primary sequence of cyclic nucleotide phosphodiesterase isoenzymes and the design of selective inhibitors. *Trends Pharmacol Sci* **11**: 150–155, 1990.
 18. Marcoz P, Prigent AF, Lagarde M and Nemoz G, Modulation of rat thymocyte proliferative response through the inhibition of different cyclic nucleotide phosphodiesterase isoforms by means of selective inhibitors and cGMP-elevating agents. *Mol Pharmacol* **44**: 1027–1035, 1993.
 19. Banner KH and Page CP, Effect of type III and IV isoenzyme selective phosphodiesterase (PDE) inhibitors on proliferation of human peripheral blood mononuclear cells. *Br J Pharmacol* **113**: 89P, 1994.
 20. Takahashi S, Oida K, Fujiwara R, Maeda H, Hayashi S, Takai H, Tamai T, Nakai T and Miyabo S, Effect of cilostazol, a cyclic AMP phosphodiesterase inhibitor, on the proliferation of rat aortic smooth muscle cells in culture. *J Cardiovasc Pharmacol* **20**: 900–906, 1992.
 21. Souness JE, Hassall GA and Parrott DP, Inhibition of pig aortic smooth muscle cell DNA synthesis by selective type III and type IV cyclic AMP phosphodiesterase inhibitors. *Biochem Pharmacol* **44**: 857–866, 1992.
 22. Pan X, Arauz E, Krzanowski JJ, Fitzpatrick DF and Polson JB, Synergistic interactions between selective pharmacological inhibitors of phosphodiesterase isozyme families PDE III and PDE IV to attenuate proliferation of rat vascular smooth muscle cells. *Biochem Pharmacol* **48**: 827–835, 1994.
 23. Dress M, Zimmermann R and Eisenbrand G, 3',5'-cyclic nucleotide phosphodiesterase in tumor cells as potential target for tumor growth inhibition. *Cancer Res* **53**: 3058–3061, 1993.
 24. Greenberg SM, Rosenthal DS, Greeley TA, Tantravahi R and Handin RI, Characterization of a new megakaryocytic cell line: The Dami cell. *Blood* **72**: 1968–1977, 1988.
 25. Decker T and Lohmann-Matthes ML, A quick and simple method for the quantification of lactate dehydrogenase release in measurements of cellular cytotoxicity and tumor necrosis factor (TNF) activity. *J Immunol Methods* **115**: 61–69, 1988.
 26. Lowry OH, Rosebrough NJ, Farr AL and Randall RJ, Protein measurement with the Folin phenol reagent. *J Biol Chem* **193**: 365–375, 1951.
 27. Thompson WJ, Terasaki W, Epstein PM and Strada SJ, Assay of cyclic nucleotide phosphodiesterase and resolution of multiple molecular forms of the enzyme. *Adv Cyclic Nucl Res* **10**: 69–92, 1979.
 28. Cook SJ, Archer K, Martin A, Buchheit KH, Fozard JR, Müller T, Miller AJ, Elliot KRF, Foster RW and Small RC, Further analysis of the mechanisms underlying the tracheal relaxant action of SCA40. *Br J Pharmacol* **114**: 143–151, 1995.
 29. Tallarida RJ and Murray RB, *Manual of Pharmacologic Calculation with Computer Programs*. Springer Verlag, New York, 1981.
 30. Viswanadhan VN, Ghose AK, Revankar GR and Robins RK, Atomic physicochemical parameters for three dimensional structure directed quantitative structure-activity relationships. 4. Additional parameters for hydrophobic and dispersive interactions and their application for an automated superposition of certain naturally occurring nucleoside antibiotics. *J Chem Infr Comput Sci* **29**: 163–172, 1989.
 31. Ruppert D and Weithmann KU, HL 725, an extremely potent inhibitor of platelet phosphodiesterase induced platelet aggregation *in vitro*. *Life Sci* **31**, 2037–2043, 1982.
 32. Sheppard H, Wiggan G and Tsien WH, Structure activity relationships for inhibitors of phosphodiesterase from erythrocytes and other tissues. *Adv Cyclic Nucleotide Res* **1**: 103–112, 1972.
 33. Hagiwara M, Endo T, Kanayama T and Hidaka H, Effect of 1-(3-chloroanilino)-4-phenylphthalazine (MY-5445), a specific inhibitor of cyclic GMP phosphodiesterase, on human platelet aggregation. *J Pharmacol Exp Ther* **228**: 467–471, 1984.
 34. Harrison SA, Reifsnnyder DA, Gallis B, Cadd GG and Beavo JA, Isolation and characterisation of bovine cardiac muscle cGMP-inhibited phosphodiesterase: a receptor for new cardiotonic drugs. *Mol Pharmacol* **29**: 506–514, 1986.
 35. Schneider HH, Schmiechen R, Brezinski M and Seidler J, Stereospecific binding of the antidepressant rolipram to brain protein structures. *Eur J Pharmacol* **127**: 105–115, 1986.
 36. Gillespie PG and Beavo JA, Inhibition and stimulation of photoreceptor phosphodiesterases by dipyrindamole and M&B 22,948. *Mol Pharmacol* **36**: 773–781, 1989.
 37. Beavo JA, Cyclic nucleotide phosphodiesterase: Functional implications of multiple isoforms. *Physiol Rev* **75**: 725–748, 1995.
 38. Tomes C, Rossi S and Moreno S, Isobutylmethylxanthine and other classical cyclic nucleotide phosphodiesterase inhibitors affect cAMP-dependent protein kinase activity. *Cell Signalling* **5**: 615–621, 1993.
 39. Parsons WJ, Ramkumar V and Stiles GL, Isobutylmethylxanthine stimulates adenylate cyclase by blocking the inhibitory regulatory protein, Gi. *Mol Pharmacol* **34**: 37–41, 1988.
 40. Manganiello VC, Murata T, Taira M, Belfrage P and Degerman E, Perspectives in biochemistry and biophysics—diversity in cyclic nucleotide phosphodiesterase isoenzyme families. *Arch Biochem Biophys* **322**: 1–13, 1995.
 41. Vincentini LM, Ambrosini A, Di virgilio F, Meldolesi J and Pozzan T, Activation of muscarinic receptors in PC12 cells. Correlation between cytosolic Ca²⁺ rise and phosphoinositide hydrolysis. *Biochem J* **234**: 555–562, 1986.
 42. Pyne NJ and Burns F, Lung phosphodiesterase isoenzymes. *Agents Actions Suppl* **43**: 35–49, 1993.
 43. Burns F and Pyne NJ, Interaction of the catalytic subunit of protein kinase A with the lung type V cyclic GMP phosphodiesterase: modulation of non-catalytic binding sites. *Biochem Biophys Res Commun* **189**: 1389–1396, 1992.